

Dissect Aid®

Suggested Methodology

Types of Specimens best suited for Dissect Aid:

- a. Lymph node embedded sections
- b. Fatty sections
- c. Kidney
- d. Breast
- e. Colon
- f. Other similar tissue samples

When to use Dissect Aid:

Dissect Aid should be used on fresh tissue section during or just after gross pathology. You may wish to remove some lymph nodes for immediate fixation and processing before gross pathology.

Fixation Time:

The smaller the section, the faster the fixation. Although radical cases may be fixed whole, they will fix more rapidly if small cuts are made into the tissue or if the tissue has been breadloafed. Place the tissue in a container and cover with Dissect Aid. We recommend that 10 times the volume of fixative to tissue ratio be used. Specimens that have been breadloafed or sliced through should fix within 3-4 hours. Entire radical sections (such as radical breast or colon) may be fixed overnight.

Post Fixation:

After fixation, sections should be rinsed thoroughly in running water for at least 1 to 2 minutes. Processing is routine once fixation has occurred. Specimens may be transferred to 10% Neutral Buffered Formalin (or other suitable fixative) for long term storage.

Results:

Dissect Aid contains alcohol, water, formaldehyde and glacial acetic acid. It will fix just like 10% Neutral Buffered Formalin. It will also firm up the ancillary fat in lymph node embedded tissue. Lastly, it will turn all lymph nodes a pale white so that they stand out in the surrounding tissue mass.

If you have any questions or would like to speak with a colleague who uses Dissect Aid, please call 1-800-428-5856 or email us at Sales@decal-bone.com

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